

Los Alamos HIV Immunology Database Overview

Bette Korber

**Summer School on Quantitative Systems Immunology,
Boston University
June 10-15, 2013**

HIV Immunology database PI: Karina Yusim

slides available as PDF documents:

<http://www.hiv.lanl.gov/content/sequence/HIV/HIVWORKSHOP/index.html>

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Overview

Immunology database introduction

Epitope maps and epitope summary tables

T-cell epitope search

T-cell epitope variants

Antibody search

Features of most broadly neutralizing antibodies (antibody “A-list”)

Neutralizing antibody resources: contexts and features – new

QuickAlign – Align an epitope to the HIV database alignments

N-glycosite – finds N-linked glycosylation sites

ELF – epitope location finder

Peptgen – list peptides for reagent development

HeatMap

HIV genome browser tools – coming soon

Mosaic Vaccine Maker, Epicover, and Posicover

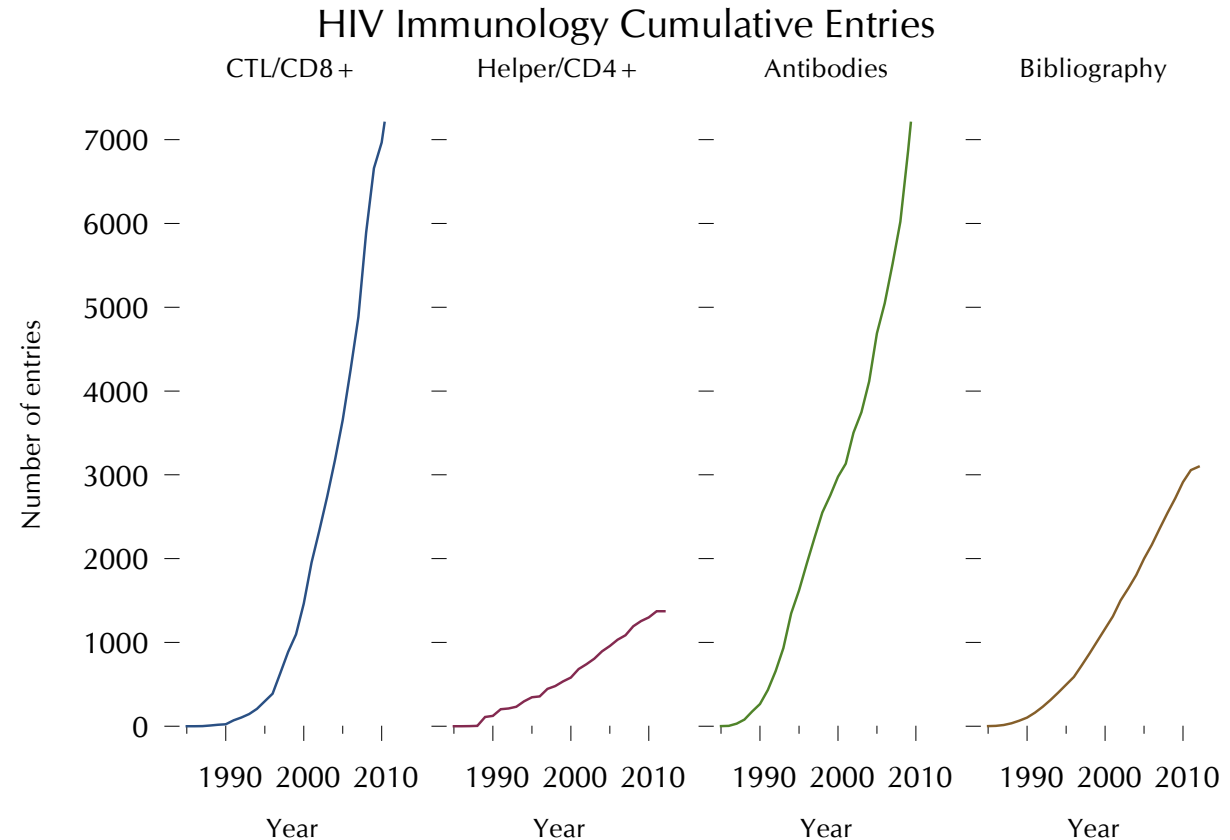
- generate candidate vaccines*
- estimate epitope coverage*
- determine regional epitope coverage*

Immunology Database Content

- Incorporates published HIV T cell (CTL, T-helper) epitope and Antibody information (emphasis on monoclonals)
- Key information regarding what is learned about epitopes and mAbs in each paper is included
- Types of data recorded:
 - Epitope sequence and location: HXB2 numbering, subtype
 - Natural infection or vaccine
 - Host HLA or MHC
 - Ab isotype, binding region, species
 - Notes summarize main findings

Immunology Database Statistics

- Contents: data from 1985 through 2013
 - 7441 CTL entries
 - 1315 T-helper entries
 - 2386 Ab entries
 - 3090 published citations



- Usage:
 - ~ 70% papers entered in CTL epitope database use HIV Immunology Database resources

HIV Molecular Immunology Database

The HIV Molecular Immunology Database is an annotated, searchable collection of HIV-1 cytotoxic and helper T-cell epitopes and antibody binding sites.

Search the Molecular Immunology Database

- [CTL/CD8+ Search](#)
- [T Helper/CD4+ Search](#)
- [Antibody Search](#)
- [Search Help](#)

Database Products

- [All Database products and publications](#)
- [Epitope maps](#)
- [Epitope summary tables](#)
- [Epitope alignments](#)
- [Epitope variants and escape mutations](#)
- [The HIV Molecular Immunology Compendium](#)
- [About the HIV Molecular Immunology Database](#)
- [How to cite this database](#)

Tools and Data Sets

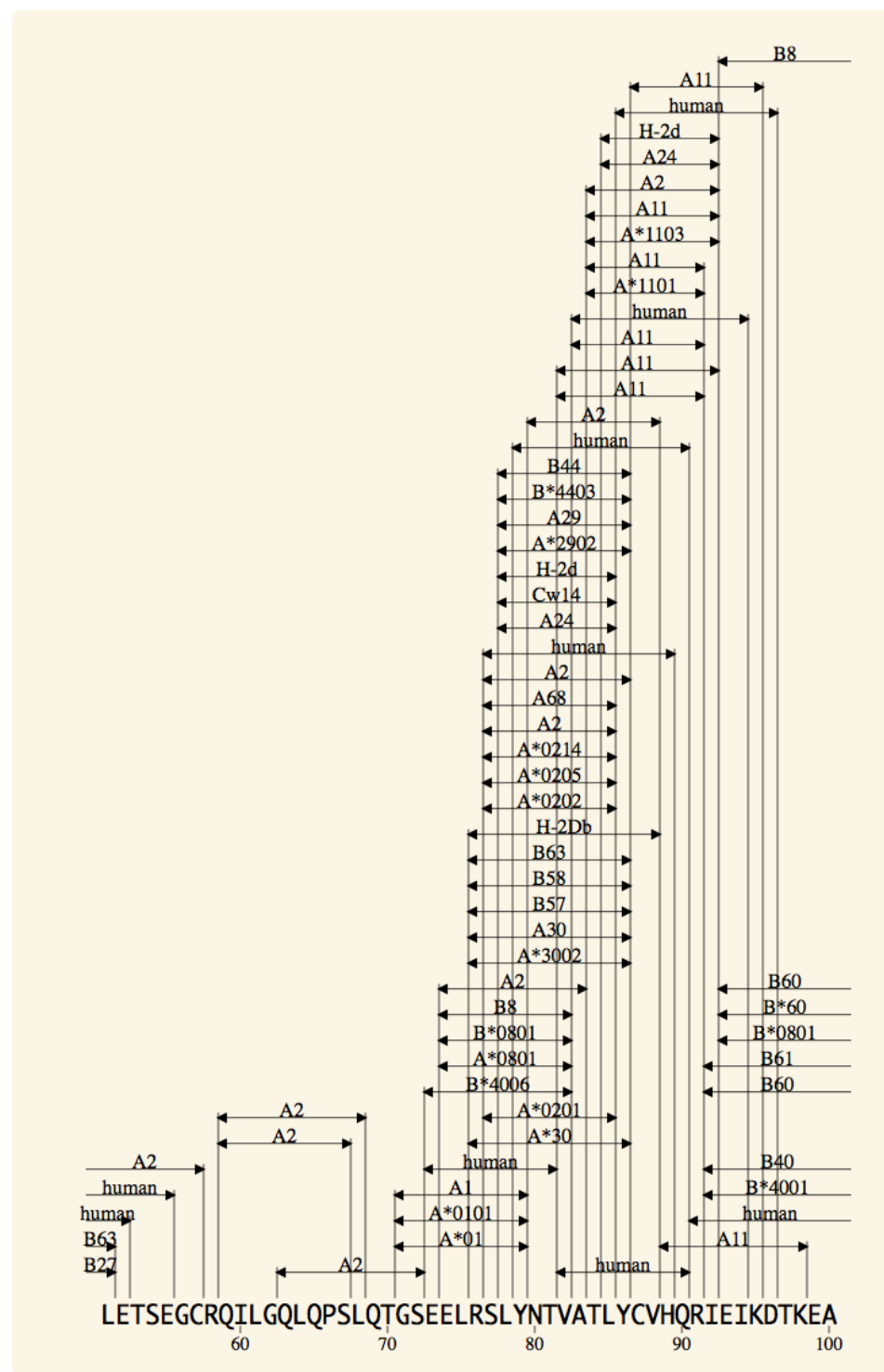
- [Tools & Links](#) for immunologists
- [HIV "A list" CTL/CD8+ Epitopes \(PDF\)](#) review article summarizing the best-characterized HIV epitopes
- [SIV Epitopes \(PDF\)](#) review article summarizing known SIV epitopes
- [Identifying HLA-Associated Polymorphisms in HIV-1 \(PDF\)](#) review article summarizing HIV polymorphism associated with escape mutations. Also a [table of polymorphisms](#).
- [HLATEM](#) HLA Typing and Epitope Mapping Data Sets
- [Standardized Assessments of Neutralizing Antibodies for HIV/AIDS Vaccine Development](#) Assay protocols from Duke Central Reference Laboratory

Immunology Database Products

- Epitope maps (species/HLA for T cell epitopes; species/MAb name for Ab)
- Epitope summary tables:
 - All CTL and Helper epitopes and Ab binding sites
 - Variants of CTL epitopes
 - Review of HIV-1 HLA-associated polymorphisms, with a link to a table of polymorphisms, by Zabrina Brumme *et al.* (Under “Tools and Data Sets”).
 - Christian Brander keeps an “A list” of HIV CD8+ T-cell epitopes – experimentally validated optimal epitopes with known HLA presenting molecules, will be updated soon
 - “B list” – a comprehensive list of all unique epitopes in the database (unknown HLA, boundaries not fully defined...)
 - All antibodies organized by protein and binding region
 - **New:** Neutralizing Ab resources: Antibody A-list, Antibody contexts and features
- Tools for immunologists
- Yearly HIV Molecular Immunology Compendium

p17 CTL/CD8+ Epitope Map

- Epitopes up to 14 aa long are mapped on HXB2
- HXB2 sequence may differ
- Epitopes with identical boundaries and HLA fields are included in the maps only once
- The epitope maps are interactive!



CTL/CD8+ Epitope Summary (B-list)

- List of all epitopes up to 21 aa long
- Unlike epitope maps that show epitope locations, here each epitope sequence is shown

Epitope	Protein	HXB2 Location	Subtype	Species	HLA
MGARASVLSG	p17	1-10	CRF01_AE	human	
ASVLSGGEL	p17	5-13	B	human	
ASILRGGKLDK	p17	5-15	C	human	
SVLSGGQLDR	p17	6-15	B	human	A11
LSGGELDRWEK	p17	8-18		macaque	
GELDRWEKI	p17	11-19	B	human	B*4002, B40
QLDRWEKI	p17	11-19	B	human	
GKLDSWEKIRLR	p17	11-22	A, CRF01_AE, CRF02_AG	human	
GKLDAWEKIRLR	p17	11-22	CRF01_AE	human	
ELDRWEKIRL	p17	12-21	B, C	human	B63
EKIRLRPGGKKYKL	p17	17-31		human	B27, B7
KIRLRPGGK	p17	18-26	A, A1, B, CRF01_AE	human, transgenic mouse	A*0301, A11, A3, B27, B7
KIRLRPGGKK	p17	18-27	B, C, multiple	human	A*0301, A11, A3, B27
KIRLRPGGKKKYKL	p17	18-31		human	A3, B62

Best-defined CTL/CD8+ Epitope Summary (A-list)

- Selective list of best defined epitopes as described by Christian Brander and colleagues

Epitope	Protein	HXB2 Location	Subtype	Species	HLA
GELDRWEKI	p17	11-19		human	B*4002
KIRLRPGGK	p17	18-26		human	A*0301
IRLRPGGKK	p17	19-27	B	human	B*2705
RLRPGGKKK	p17	20-28		human	A*0301
RLRPGGKKKY	p17	20-29	B	human	A*0301
GGKKKYKLK	p17	24-32	B	human	B*0801
KYKLKHIVW	p17	28-36	B	human	A*2402
HLVWASREL	p17	33-41		human	Cw*0804
LVWASRELERF	p17	34-44		human	A30
WASRELERF	p17	36-44	B	human	B*3501
ELRSLYNTV	p17	74-82		human	B*0801
RSLYNTVATLY	p17	76-86	B	human	A*3002, B58, B63
SLYNTVATL	p17	77-85	B	human	A*0201, A*0202, A*0205
LYNTVATL	p17	78-85		human	Cw14
LYNTVATLY	p17	78-86		human	A*2902, B*4403
TLYCVHQK	p17	84-91		human	A*1101
IEIKDTKEAL	p17	92-101		human	B*4001
NSSKVSQNY	p17	124-132	B	human	B*3501

Immunology Database: Search

■ T Cells

- ☐ Cytotoxic T Lymphocytes (CTL)
- ☐ Helper T Lymphocytes (T-helper)
- ☐ Organization is identical for CTL and T-helper
- ☐ One reference per entry, epitope/HLA combinations are often repeated

■ B Cells (Antibodies)

- ☐ One entry for each monoclonal antibodies
- ☐ Many references per entry (> 400 for some well studied MAbs)

CTL/CD8+ T-cell Search

- Can search by HIV protein, Epitope Sequence, Subtype, Immunogen, Vaccine Details, Species, presenting MHC/HLA, Author, Country, Keywords
- Can now search on epitope location and find fuzzy matches, overlaps and embedded epitopes
- Search example:
 - SLYNTVATL – 254 entries
 - To narrow the search use keyword “escape” – 32 entries
- Additional information provided in the entry:
 - Location, Donor MHC/HLA, experimental methods, Notes
 - CTL epitope variants if studied in the paper
 - Link to all entries for a reference
 - PubMed links to papers
 - Link to Epitope Maps
 - Link to Epitope Alignment (Extracted from HIV-sequence database, includes subtype, country and year of sampling)

CTL/CD8+ T-cell Search

Search for

Epitope: ISPRTLNAW

First Author: Pillay

HIV protein	Proteins with defined epitopes - ALL - p17 p17-p24 p24 p24-p2p7p1p6	Proteins with undefined epitopes - ALL - Gag Gag/Pol Pol Vif
HXB2 location	<input type="text"/> - <input type="text"/>	<input type="button" value="Results overlap with query location"/>
Epitope	<input type="text" value="ISPRTLNAW"/>	<input type="button" value="Results contain query sequence"/>
Epitope name	<input type="text"/>	
Record number	<input type="text"/>	
Subtype	- ALL -	
Immunogen	- ALL - computer prediction HIV-1 and GBV-C co-infection HIV-1 and HCV co-infection HIV-1 exposed seronegative HIV-1 infected monocyte-derived HIV-1 infection	
Vaccine details	Vaccine type Vaccine strain if Immunogen is Vaccine Vaccine component Adjuvant	- ALL - - ALL - - ALL - - ALL -
Species	- ALL -	
MHC/HLA	- ALL - A*01 A*0101 A*02 A*0201 A*02.01 A*020101	
Author	<input type="text" value="Pillay"/>	<input checked="" type="checkbox"/> First <input type="checkbox"/> Last
Country	- ALL -	
Keywords	- ALL - acute/early infection adjuvant comparison antagonism antibody binding site definition and exposure assay development, comparison, standardization, improvement autologous responses	
Note	<input type="text"/>	

 Click for [Search Help](#)

Search CTL/CD8+ T-Cell Epitope Database

Found 1 matching record:

Displaying record number 53832

HXB2 Location	p24(15-23)
Author Location	Gag(147-155)
Epitope	ISPRTLNAW
Subtype	C
Species (MHC/HLA)	human(B57)
Immunogen	HIV-1 infection
Donor MHC/HLA	A*3001, A*66, B*4201, B*5802, Cw*0602, Cw*1701; A*66, A*68, B*57, B*5802, Cw*0602, Cw*0701
Country	South Africa
Experimental methods	CD8 T-cell Elispot - IFN γ
Keywords	epitope processing, responses in children, mother-to-infant transmission, escape, acute/early infection

Notes

- HIV-specific CTLs in infants were shown to be able to select for viral escape variants early in life, despite a lack of escape with the same CTL specificity in the mother. Infant CTL responses may be compromised by transmission of escape variants that arose in the mother and also those that arose in the father, if the father was the source of the mother's infection.
- ISPRTLNAW is the C consensus form of the epitope and was the autologous form in the mother, and was transmitted to her infant. By 33 weeks a new dominant form of the epitope had emerged in the infant, mSPRTLNAW, and two additional variants had arisen, one with a substitution proximal to the epitope, pISPRTLNAW, and ISPRTLNAW.

References

Pillay2005 Thillagavathie Pillay, Hua-Tang Zhang, Jan W. Drijfhout, Nicola Robinson, Helen Brown, Munira Khan, Jagadesa Moodley, Miriam Adhikari, Katja Pfafferott, Margaret E. Feeney, Anne St. John, Edward C. Holmes, Hoosen M. Coovadia, Paul Klenerman, Philip J. R. Goulder. and Rodnev E. Phillips. Unique Acquisition of Cytotoxic T-Lymphocyte Escape Mutants in Infant Human Immunodeficiency

Epitope Map

[p24 Epitope Map](#)

[Epitope Alignment](#)

[Show epitope variants](#)

Epitope alignment to Database sequences

Variant details with annotator's notes

Variants details

HXB2 Location	p24(15-23)	p24 Epitope Map
Epitope	ISPRTLNAW	Epitope Alignment
Variants	mSPRTLNAW	escape documented in this paper
	lSPRTLNAW	diminished response
	pI lSPRTLNAW	not determined
Species (MHC/HLA)	human(B57)	

Can go back to epitope entry

Variant Details

Showing all 3 variants.

Variant ID.	1413
Epitope Seq.	ISPRTLNAW
Variant Seq.	mSPRTLNAW
Mutations	I/M
Epitope Location	I1M
HXB2 Location	I15M
Mutation Type	E: escape documented in this paper
Method	CD8 T-cell Elispot - IFNy, Sequence
Note	This is de novo variant seen in infant by week 33 of age. The index peptide was recognized, but not the variant.

Mutation type

Note describing why the variant was designated particular mutation type

Variant ID.	1414
Epitope Seq.	ISPRTLNAW
Variant Seq.	lSPRTLNAW
Mutations	I/L
Epitope Location	I1L
HXB2 Location	I15L
Mutation Type	DR: diminished response
Method	CD8 T-cell Elispot - IFNy, Sequence

Summary table of ~ 2800 variants

CTL/CD8+ Epitope Variant Details

Download CTL/CD8+ epitope variant details as [CSV](#) or [XLS](#) files.

[List of Mutation types](#)

Data last updated at 2013-01-25 11:58:16-07

Epitope ID	Epitope Name	Variant ID	Subtype	Epitope Subtype	Variant Subtype	Protein	HXB2 start	HXB2 end	HLA	Epitope	Variant Epitope	Mutation (epitope)	Mutation (protein)	Mutation Type Code	Mutation Type Description	Methods	Note	References
54532	AI14	1016	B	B	A, M-group	p17	5	19		ASVLSGGELDRWEKI	ASVLSGGKLDaWEKI	R11A, E8K	R15A, E12K	SNSF	subtype-specific non-susceptible form	CD8 T-cell Elispot - IFN γ	No cross-recognition of this variant was seen across clades or intra-clade central sequences.	Malhotra2007
54532	AI14	1017	B	B	C	p17	5	19		ASVLSGGELDRWEKI	ASILrGGKLDKWEKI	R11K, V3I, S5R, E8K	R15K, V7I, S9R, E12K	SNSF	subtype-specific non-susceptible form	CD8 T-cell Elispot - IFN γ	No cross-recognition of this variant was seen across clades or intra-clade central sequences.	Malhotra2007
54532	AI14	1018	B	B	B	p17	5	19		ASVLSGGELDRWEKI	ASVLSGGKLDKWEKI	R11K, E8K	R15K, E12K	SNSF	subtype-specific non-susceptible form	CD8 T-cell Elispot - IFN γ	No cross-recognition of this variant was seen across clades or intra-clade central sequences.	Malhotra2007
54532	AI14	1019	B	B	B	p17	5	19		ASVLSGGELDRWEKI	ASVLSGGELDKWEKI	R11K	R15K	SNSF	subtype-specific non-susceptible form	CD8 T-cell Elispot - IFN γ	No cross-recognition of this variant was seen across clades or intra-clade central sequences.	Malhotra2007
53591	Gag 1.2	54		B	CRF02_AG	p17	8	18		LSGGELDRWEK	LSGGKLDaWEK	E5K, R8A	E12K, R15A	SNSF	subtype-specific non-susceptible form	Intracellular cytokine staining, T-cell Elispot	CRF02 form, LSGGKLDaWEK, does not cross-react with the B clade LSGGELDRWEK elicited response.	Amara2005a
53844	GI9	1569	B			p17	11	19	B40	GELDRWEKI	GELDRWkKI	E7K	E17K	DR, LE	diminished response, literature escape	CD8 T-cell Elispot - IFN γ , Sequence	This variant from the HXB2 sequence was present in the restricting HLA-B40-carrying mother, M-1002, but was never detected in her non-HLA-B40-carrying infant, P-1031. Decreased recognition of the E17K variant relative to the index epitope was seen in the mother.	Sanchez-Merino2005
56027	GI9(p17)	1903	B	B	B	p17	11	19		GQLDRWEKI	GeLDRWEKI	Q2E	Q12E	ND	not determined	CD8 T-cell Elispot - IFN γ , Sequence	This Asian B Clade optimal epitope differs from the consensus B at one position. It is predicted to be HLA-B40 restricted. Experimentally, B clade consensus peptide was used to challenge CTL response in subjects commonly carrying the Asian B-type epitope.	Zhai2008
55632		11	A, CRF02_AG, CRF01_AE	A, AG	AE	p17	11	22		GKLDSEWKIRLR	GKLDaWEKIRLR	S5A	S15A	SSF	subtype-specific susceptible form	CD8 T-cell Elispot - IFN γ	1 subject responded to peptide GKLDSEWKIRLR from subtypes CRF02_AG and A and to peptide GKLDaWEKIRLR from subtype CRF01_AE.	Aldoo2008
54629	GAG-03	1957	B	B	C	p17	17	34		EKIRLRPGGKKYRLKHL	EKIRLRPGGKKhYmLKHL	K12H, R14M	K28H, R30M	SSF	subtype-specific susceptible form	CD8 T-cell Elispot - IFN γ , Sequence	This Clade C consensus synthetic peptide variant from an immunodominant region, differs from the immunodominant Clade B consensus at 2 amino acids (11,1%) and both were recognized by subtype-B-infected subjects.	Zhao2007
53201	KK9	31	B			p17	18	26	A3	KIRLRPGGK	KIRLRPGGq	K9Q	K26Q	E, P	escape documented in this paper, processing	CD8 T-cell Elispot - IFN γ , Flow cytometric T-cell cytokine assay	Variant inhibits processing, resulting in rapid decline in the KK9 specific CD8+ T-cell response.	Allen2004
55770	KK9	153	B			p17	18	26	A3	KIRLRPGGK	KIRLRPGGr	K9R	K26R	SF	susceptible form	Flow cytometric T-cell cytokine assay	KIRLRPGGK was recognized by 3 patients. The autologous sequence in one patient was KIRLRPGGr which induced high frequency response.	Daucher2008
55233		790	B, CRF01_AE		B	p17	18	26	A3	KIRLRPGGK	KIRLRPGGr	K9R	K26R	IE	inferred escape	Sequence	Patient was superinfected with three strains, B1, B2 and CRF01_AE. This variant developed in B1 to include 42% of the viruses within 4 years.	Kozaczynska2007
This variant was seen in Donnot and																		

Under database Products

(<http://www.hiv.lanl.gov/content/immunology/index.html>)

Go to “Epitope variants and escape mutations”

Antibody Search

- Can search by
 - HIV protein, Epitope Sequence, Subtype, Immunogen, Vaccine Details, Species, presenting MHC/HLA, Author, Country, Keywords
 - MAb ID (Ab lists by name and by binding type are provided)
 - Ab type (by binding site, for example binding to similar region like V3 or near a common functional domain like CD4 binding site CD4Bs)
 - Isotype
- Search examples:
 - 2F5 – 1 record with 463 references
 - Ab type: gp120 CD4BS – 200 records

Antibody Search

HIV protein	Proteins with defined epitopes - ALL - p17 p17-p24 p24 p24-p2p7p1p6	Proteins with undefined epitopes - ALL - p24 Gag RT Pol
HXB2 location	<input type="text"/> - <input type="text"/>	Results overlap with query location
Epitope	<input type="text"/>	Results contain query sequence
Record number	<input type="text"/>	
MAb ID	<input type="text"/>	(List by name) (List by type)
Subtype	- ALL -	
Immunogen	- ALL - anti-idiotypic autoimmune disease HIV-1 exposed seronegative HIV-1 infection HIV-2 infection in vitro stimulation or selection	
Vaccine details if Immunogen is Vaccine	Vaccine type Vaccine strain Vaccine component Adjuvant	- ALL - - ALL - - ALL - - ALL -
Ab Type	- ALL - C-domain C-HR C-term Env oligomer flap region gp120 adjacent to CD4BS	
Species	- ALL -	
Isotype	- ALL - IgA IgA1 IgA2 IgA22a IgE IgG	
Author	<input type="text"/>	Search only for <input type="checkbox"/> First <input type="checkbox"/> Last author <input checked="" type="radio"/> Show only this author's references <input type="radio"/> Show all references
Country	- ALL -	
Keywords	- ALL - acute/early infection ADCC adjuvant comparison antibody binding site definition and exposure antibody generation antibody interactions	<input checked="" type="radio"/> Show only notes containing selected keyword(s) <input type="radio"/> Show all notes
Note	<input type="text"/>	<input checked="" type="radio"/> Show only notes matching this text <input type="radio"/> Show all notes

Can search by HXB2 location,
Find overlaps, fuzzy matches
Embedded epitopes

Can show only notes and
references containing
selected keywords or user's
text (as apposed to showing
matching Ab entries with all
notes)

Search

Reset

Click for [Search Help](#)

Antibody Search

Found 1 matching record:

Displaying record number 815

MAb ID	2F5 (IAM 2F5, IAM-41-2F5, IAM2F5, c2F5)	gp160 Epitope Map
HXB2 Location	gp160(662-667)	
Author Location	gp41(662-667 BH10)	
Research Contact	Hermann Katinger, Institute of Applied Microbiology, Vienna, or Polymun Scientific Inc., Vienna, Austria	
Epitope	ELDKWA	Epitope Alignment
Ab Type	gp41 adjacent to cluster II, C-term, gp41 MPER (membrane proximal external region)	
Neutralizing	L P	
Species (Isotype)	human(IgG3κ)	
Immunogen	HIV-1 infection	
Keywords	acute/early infection, adjuvant comparison, anti-idiotypic, antibody binding site definition and exposure, antibody generation, antibody interactions, antibody sequence variable domain, assay development, standardization and improvement, autoantibody or autoimmunity, autologous responses, binding affinity, brain/CSF, co-receptor, complement, dendritic cells, drug resistance, enhancing activity, escape, genital and mucosal immunity, HAART, ART, HIV exposed persistently seronegative (HEPS), immunoprophylaxis, immunotherapy, immunotoxin, isotype switch, kinetics, mimics, mimotopes, mother-to-infant transmission, neutralization, rate of progression, responses in children, review, SIV, structure, subtype comparisons, supervised treatment interruptions (STI), therapeutic vaccine, vaccine antigen design, vaccine-induced immune responses, variant cross-recognition or cross-neutralization, viral fitness and reversion	

Notes

- 2F5: 2F5 neutralized infection of PBLs with various HIV-1 strains with high potency. However, 2F5 did not inhibit transcytosis of cell-free or cell-associated virus across a monolayer of epithelial cells. A mixture of 13 MAbs directed to well-defined epitopes of the HIV-1 envelope, including 2F5, did not inhibit HIV-1 transcytosis, indicating that envelope epitopes involved in neutralization are not involved in mediating HIV-1 transcytosis. When the mixture of 13 MAbs and HIV-1 was incubated with polyclonal anti-human γ chain, the transcytosis was partially inhibited, indicating that agglutination of viral particles at the apical surface of cells may be critical for HIV transcytosis inhibition by HIV-specific Abs. [Chomont2008](#) (neutralization)
- 2F5: The lipid binding properties of 2F5, and the similarity to binding properties of anti-lipid mAbs, are discussed. Potential role of liposomes containing lipid A for induction of NABs to lipids of HIV-1 is reviewed. [Alving2008](#) (autoantibody or autoimmunity, review)
- 2F5: A reference panel of recently transmitted Tier 2 HIV-1 subtype B envelope viruses was developed representing a broad spectrum of genetic diversity and neutralization sensitivity. The panel includes viruses derived from male-to-male, female-to-male, and male-to-female sexual transmissions, and CCR5 as well as CXCR4 using viruses. The envelopes displayed varying degrees of neutralization sensitivity to 2F5, with 14 of 19 envelopes sensitive to neutralization by this Ab. [Schweighardt2007](#) (assay development, standardization and improvement, neutralization)
- 2F5: This review summarizes data on possible vaccine targets for elicitation of neutralizing Abs and discusses whether it is more practical to design

Neutralizing Antibody Resources

Summary table

A table presenting most broadly neutralizing HIV-1 antibodies, with links to papers, Ab sequences and structure, notes on breadth of neutralization, where to find Ab contacts or key residues, and heavy and light chain composition.

- [Best neutralizing antibodies](#)

Search interface

This interface allows you to search for exact coordinates of important neutralizing antibody binding sites and other HIV-1 Env features.

- [Neutralizing antibody contexts and features](#)

Spreadsheet

A summary of the information from the search interface above, presented in a single .xls spreadsheet. Each row of the table corresponds to one residue of HIV-1 Env, and each column represents a protein feature or set of known binding residues of broadly neutralizing antibodies. Loops and other features of Env are shown in the first 3 columns on the left. The entropy (sequence variability) of each residue is presented numerically and color coded. Abbreviated references are listed under each column heading, and full references are on the second page of the Excel file.

- [Spreadsheet of neutralizing antibody contexts and features](#) (.xls)

Summary of Best Neutralizing Antibodies

Download summary of best neutralizing antibodies as [CSV](#) or [XLS](#) files.

This is a list of most broadly neutralizing HIV antibodies, with links to papers, Ab sequences and structure, notes on breadth of neutralization, Ab contact or key residues and heavy and light chain composition. Note: this is a work in progress, so not all relevant papers and antibodies are listed.

Mab	Binding site	Author Journal Pmid	First paper	Breadth of neutralization with IC50<50 µg/ml	Breadth of neutralization with IC80 or IC90<50 µg/ml	Structure, PDB ID	Ab sequence	Heavy chain	Light chain	Germline Ab sequence	Ab binding affinity	Listings of antibody contact or key residues
VRC01	CD4bs	Wu2010 Science 20616233	YES	91% of 190 isolates, representing major HIV-1 clades	86% of 190 isolates, representing major HIV-1 clades, with IC80		GI:294875838 -- heavy chain variable region GI:294875848 -- light chain variable region	V: IGHV1-02*02 D: IGHD3-16*01 (or *02) J: IGHJ1*01 or IGHJ2*01	V: IGKV3-11*01 J: IGKJ2*01	Fig. S5	Bound strongly to RSC3 and gp120 and weakly to ΔRSC3, Fig. 2 and S4.	
		Zhou2010 Science 20616231				3NGB				Fig. S12	Figs. 5, 6, S3	Env, defined by crystal structure: Fig S1. Antibody, defined by crystal structure: Fig. S9
		Wu2011 Science 21835983								Sequence, Figs. 1, S14, S18. Phylogenetic analysis, Fig. 5, Fig. S13		Antibody, defined by crystal structure, compared to key residues of other CD4bs antibodies, Fig. S4.
		Scheid2011 Science 21764753		100% of 118 isolates representing major HIV-1 clades							Fig. 3, Table S9.	Antibody, defined by crystal structure in Zhou2010, Fig. 4, Fig. S3, and Fig. S4 provide comparisons with other CD4bs Nabs.
		Walker2011 Nature 21849977		93% of 162 isolates representing major HIV clades	89% of 162 isolates representing major HIV clades, with IC90							

<http://hiv-dev.lanl.gov:8081/content/immunology/tables/tables.html>

Epitope Summary Tables
Best Neutralizing Antibodies

Spreadsheet of Ab contexts and features

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	AA
	References are shown below each column heading. Full references are in SHEET 2 of this Excel file																										
	Regions_1: signal peptide, cleavage sites, disulfied bonds, hypervariable regions, integrin α4β7 binding, Lectin DC-sign binding, gp41 regions: Leucine/Isoleucine	Regions_2: Variable loops, gp41 regions: Kennedy epitope, GxxxG motif, Cytoplasmic tail LLP1, LLP2, LLP3; RxKR motif	Regions_3: gp120 Regions related to the CD4 binding site, gp41 regions: fusion peptide, Leucine/Isoleucine zipper, fusion domain, YXXL motif	Entropy score	HXB2 position	HXB2 AA	CH103 contact & bonding	b12 contacts	17b contacts	PG9 contacts PG9-related class: PG16, PGT141, 145, CH01-CH04	PG9 sensitivity PG9-related class: PG16, PGT141, 145, CH01-CH04	10E8	2F5	4E10	Z13												
8	Cys 131 linked to Cys 157	forms V1 loop		0.02	157	C																					
9		V2		0.30	158	S																					
0		V2		0.17	159	F																					
1		V2		0.40	160	N																					
2		V2		1.57	161	I																					
3		V2		0.29	162	S																					
4		V2		0.36	163	T																					
5		V2		1.66	164	S																					
6		V2		1.32	165	I																					
7		V2		1.13	166	R																					
8		V2		0.60	167	G																					
9		V2		0.55	168	K																					
0		V2		1.97	169	V																					
1		V2		1.46	170	Q																					
2		V2		1.24	171	K																					
3		V2		1.51	172	E																					
4		V2		1.17	173	Y																					
5		V2		0.42	174	A																					
6		V2		0.50	175	F																					
7		V2		0.17	176	F																					
8		V2		0.35	177	Y																					
9		V2		1.23	178	K																					
0	LDI/LDV tripeptide in V	V2		0.96	179	L																					
1	LDI/LDV tripeptide in V	V2		0.03	180	D																					
2	LDI/LDV tripeptide in V	V2		0.97	181	I																					
3		V2		0.69	182	I																					
4		V2		1.18	183	P																					
5		V2		1.06	184	I																					
6	V2-hypervariable	V2		1.70	185	D																					
7	V2-hypervariable	V2		1.87	186	N																					
8	V2-hypervariable	V2		1.82	187	D																					
9	V2-hypervariable	V2		1.87	188	T																					
0	V2-hypervariable	V2		1.81	189	T																					
1	V2-hypervariable	V2		1.98	190	S																					
2		V2		0.16	191	Y																					
3		V2		0.85	192	K																					
4		V2		0.14	193	L																					
5		V2		0.50	194	T																					
6		V2		0.90	195	S																					
7	Cys 126 linked to Cys 196	V2 loop end		0.04	196	C																					
8				0.11	197	N																					
9				0.46	198	T																					
0				0.09	199	S																					
1				1.30	200	V																					
2				0.59	201	I																					

Tools for Immunologists

- **Sequence Locator** Finds epitope location on the reference genome
- **QuickAlign** Aligns amino acids or nucleotides against our alignments
- **PeptGen** Generates overlapping peptides for any protein
- **PepMap** Generates peptide maps in Fasta, HTML and PDF formats
- **ELF** Epitope Location Finder
- **N-Glycosite** Finds N-linked glycosylation sites
- **Mosaic** Generates candidate vaccine protein cocktails
- **Motifscan** Scans alignments for HLA binding motifs
- **Highlighter** Highlights matches and mismatches in a set of aligned sequences
- **Heatmap** Displays and organizes neutralization or other quantitative data.
- And more ...

HIV Molecular Immunology Database: Tools & Links

Tools Produced by the Los Alamos HIV Databases

- [QuickAlign](#) Align amino acids or nucleotides against our alignments
 - Epilign and PrimAlign have been replaced by QuickAlign
- [PeptGen](#) Generate overlapping peptides for any protein
- [PepMap](#) Generate peptide maps in Fasta, HTML and PDF formats
- [Hepitope](#) Search for hopeful epitopes based on HLA enrichment
- [HLA Frequency Analysis Tools](#) Calculate HLA frequencies or HLA linkage disequilibrium in a population
- [ELF](#) Epitope location finder
- [Motif Scan](#) Scan alignments for HLA binding motifs
 - [HLA genotype/serotype dictionary](#)
 - [HLA genotype/motif dictionary](#)
 - [HLA supertype dictionaries](#)
- [HIV/SIV Sequence Locator Tool](#) Formerly the *HXB2 Numbering Engine*
- [SeqPublish](#) Produce pretty alignments for publication
- [BLAST](#) Search sequences against our annotated HIV sequences
- [ODprep/ODfit](#) Calculate antibody titers based on concentration and optical density data
- [Heatmap](#) Display a table of numbers using colors to represent the numerical values
- [Mosaic](#) Generate candidate vaccine protein cocktails
 - [Epicover](#) Epitope coverage assessment tool
 - [Posicover](#) Positional epitope coverage assessment tool
- [N-Glycosite](#) Find N-linked glycosylation sites
- [Highlighter](#) Highlight matches and mismatches in a set of aligned sequences
- [All Tools](#) List of all software and tools in both the HIV sequence and immunology databases

External Tools for Epitope Prediction

- [BIMAS HLA Peptide Binding Predictions](#) Ranks potential n-mer peptides based on a predicted half-time of dissociation to HLA class I molecules
- [SYFPEITHI Epitope Prediction](#) Predicts the binding of a given amino acid sequence to a defined HLA type
- [PAProC](#) Predicts cleavages by human and yeast 20S proteasomes
- [PREDEP](#) MHC class I epitope prediction
- [MHCpred](#) Predicts MHC/peptide or TAP/peptide IC₅₀ binding values
- [Microsoft Research Epitope Predictor](#) Computes the probability that a given n-mer is a T-cell epitope restricted to a given HLA allele

Heatmap

- A heatmap is a graphical way of displaying a table of numbers by using colors to represent the numerical values.
- This is a very nice way to display large sets immunological data (eg antibody and target data), statistically clustering “like” behaviors
- Commonly used in gene expression array data
- We have two options in the immunology database:
 - Hierarchical clustering: We use R code for this, and if you are comfortable coding in R you can better manipulate fine details
 - An in-house developed k-means clustering strategy that incorporates an error model

Heatmap Hierarchical Clustering

Purpose: A heatmap is a graphical way of displaying a table of numbers by using colors to represent the numerical values. The clustering algorithm groups related rows and/or columns together by similarity. For details see [Heatmap Hierarchical Explanation](#).

Input

Paste your data here
[\[Sample Input\]](#)

Or upload a data file

Options

Use [log transformation](#)? ☐ Natural ☒ Base10 ☐ No transformation

Use [threshold value](#)? ☒ No ☐ Below threshold value: ☐ Above threshold value:

[Cluster method](#) [Distance method](#)

Heatmap ☒ Cluster by ☐ Rows only ☐ Columns only ☒ Both ☐ None

Colors [Palette:](#) [Higher values:](#) ☒ High intensity color ☐ Low intensity color [Number of colors:](#) [Color key ranges:](#)

[Margins](#) Bottom Margin: Right Margin: [Label sizes](#) Row: Column:

See the stability of clusters using [bootstraps](#) ☐ Iterations

By ☒ [Standard Bootstrap](#) (slow but accurate)
☒ scale bar
show only labels with >= % [bootstrap support](#)
☐ [pvclust](#) (fast but approximate)

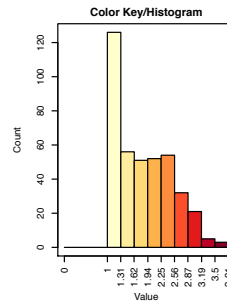
Usually use log
base 10 for
antibody data

Antibody data
is often censored

NAb sera are
dilutions, so high
values are potent

MAbs are
concentrations, so
low values are potent

K-means clustering, $k = 3$



Three clusters, red, blue, yellow

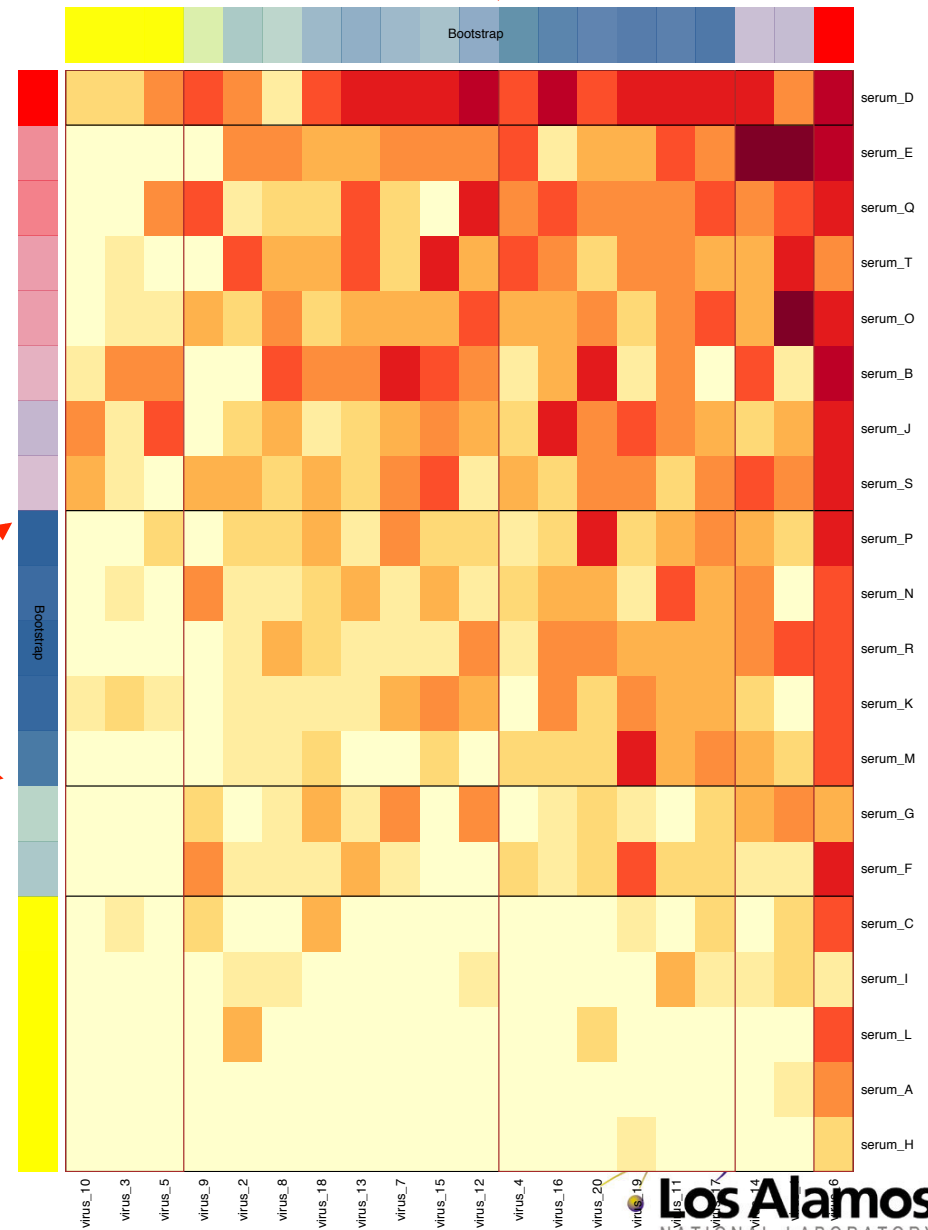
Bootstrap resampling to explore robustness of clusters given sampling limitations

Can add an error model to explore the Impact of experimental reproducibility

Robust blue cluster

Kmeans Heatmap
kmeans
of clusters = row: 3 column: 3
threshold = 80

Robust blue cluster



QuickAlign

- Generates an alignment of your HIV-1 amino acid or nucleotide sequence against our web alignments
- Can be used to align epitopes, functional domains, or any protein or nucleotide region of interest
- Calculates frequency of variants to the query sequence and summarizes both by subtype and all subtypes together
- Calculates frequency of amino acid or nucleotide by position and summarizes both by subtype and all subtypes together

QuickAlign

formerly Epilign and Primalign

Purpose: Align a desired region from our [Web alignments](#), with or without user-provided sequence(s). Details below.

Retrieve alignment(s) based on sequence

Paste your sequence(s) here

[\[Sample Input\]](#)

or upload sequence file

Browse...

-- OR leave both fields above blank, and --

Retrieve alignment(s) based on coordinates

Sequence coordinates start end

Gene/region/protein

Options

Organism ☒ HIV1 ☐ HIV2 ☐ SIV

Sequence type ☐ nucleotide ☐ protein ☒ let program decide

[Alignment type](#) to use

[Delete Gaps](#) and shift sequence toward C-terminus (protein only) ☐ yes ☒ no

Display [wide output](#) ☐ yes ☒ no

Calculate [frequency by position](#) ☒ [cut-off](#) %

Include [surrounding region](#) ☐

Submit

Reset

QuickAlign: example of output

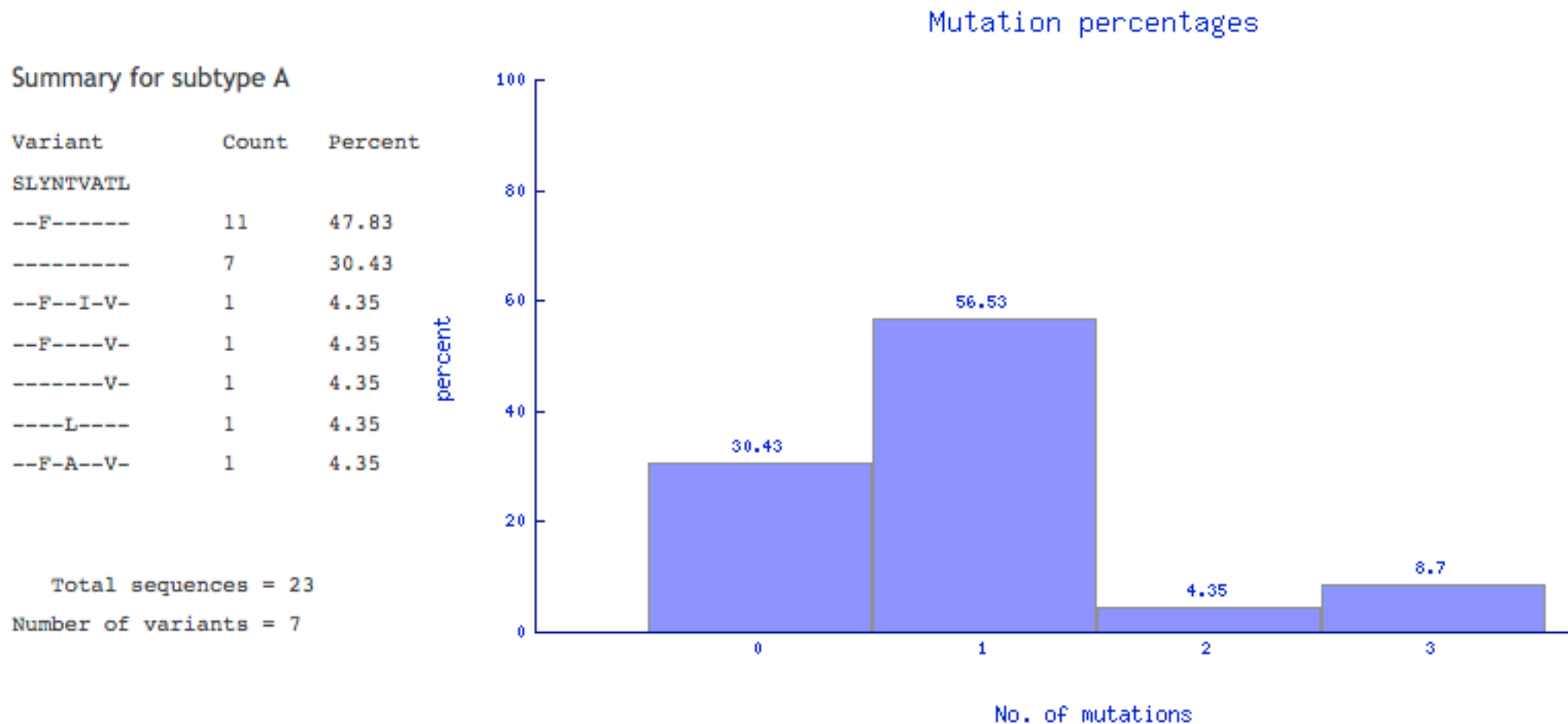
- Query peptide:
SLYNTVATL
- Sequence names
include subtype,
country and year of
sampling
- Identical sequences
are shown in red

Query:	SLYNTVATL
Query Length:	9
<u>HXB2 Location:</u>	Gag 77-85 = p17 77-85
<u>Alignment:</u>	GAG, 458 sequences

Summarize

Query	SLYNTVATL
A1.KE.86.ML170	--F-----
A1.KE.94.Q23	--F-----
A1.SE.94.SE7253	--F----V-
A1.SE.94.SE7535	-----
A1.SE.95.SE8538	-----
A1.SE.95.SE8891	-----
A1.SE.95.UGSE8131	-----
A1.TZ.97.97TZ03	--F----V-

QuickAlign: sequence variant summary



- Variant frequency summary by subtype and all subtype together

QuickAlign: Frequency by position

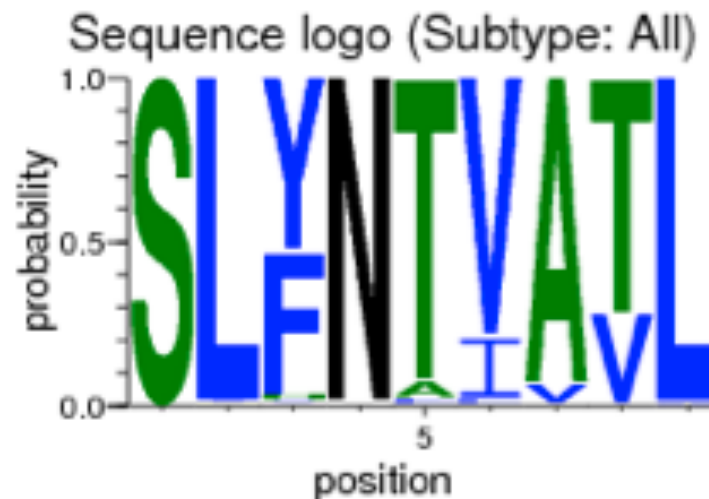
Frequency by position

[Go to top](#)

[See full raw counts](#)

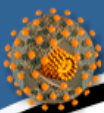
cutoff: 95%

Position	Percentage and raw count of non-gap		Non-gap/total (percentage)
1	S: 99.90% (3113)	other: 0.10% (3)	3116/3119 (100.00%)
2	L: 98.90% (3068)	other: 1.10% (34)	3102/3119 (99.55%)
3	Y: 52.71% (1633)	F: 43.77% (1356) other: 3.52% (109)	3098/3119 (99.42%)
4	N: 99.68% (3104)	other: 0.32% (10)	3114/3119 (99.94%)
5	T: 92.86% (2887)	A: 5.05% (157) other: 2.09% (65)	3109/3119 (99.78%)
6	V: 79.35% (2448)	I: 18.15% (560) other: 2.50% (77)	3085/3119 (99.01%)
7	A: 92.95% (2889)	V: 6.53% (203) other: 0.51% (16)	3108/3119 (99.74%)
8	T: 72.52% (2254)	V: 27.06% (841) other: 0.42% (13)	3108/3119 (99.74%)
9	L: 99.00% (3078)	other: 1.00% (31)	3109/3119 (99.78%)



PeptGen

- Creates maps of overlapping peptides on proteins to aid in peptide design for mapping epitopes
- Consensus sequences for all HIV subtypes for all proteins are available
- Use alignments to design comparable sets of peptides (for example, to compare clades)
- INPUT
 - Query sequence or aligned sequences
 - Desired length of peptides, peptide overlap,
 - Forbidden C- and N-terminal amino acids
- OUTPUT
 - Maps of overlapping peptides (forbidden AAs are taken into account)
 - Simplified output for ordering
 - Highlighted forbidden amino acids
 - Hydrophobicity scores for the peptides are available



PeptGen Peptide Generator

Purpose: Given an amino acid sequence, this tool generates and displays sets of overlapping peptides that can be used for peptide design and epitope mapping.

How to use: Paste amino acid sequence(s) in any valid format into the window below, or upload a file of sequences. Change the default values for the other parameters as needed and hit Submit. You can also use an [HIV consensus sequence](#) as input.

For additional details, see [PeptGen Explanation](#).

Input

Paste your input here
[\[SAMPLE INPUT - single sequence\]](#)
[\[SAMPLE INPUT - alignment\]](#)

or upload your file

If you are submitting [an alignment](#) check this box ☐

Options

Make peptides of length <input type="text" value="18"/>	C-terminal forbidden amino acids <input type="text"/>
Overlap peptides by <input type="text" value="10"/>	N-terminal forbidden amino acids <input type="text"/>
Shorten by <input type="text" value="3"/>	Apply proline rule <input checked="" type="radio"/> Yes <input type="radio"/> No
Lengthen by <input type="text" value="2"/>	Calculate hydropathy <input checked="" type="radio"/> Yes <input type="radio"/> No
	Color amino acids <input checked="" type="radio"/> Yes <input type="radio"/> No

Output

Produce [simple output](#) ☐

Duplicate peptides ☒ flag ☐ remove

Gaps ☒ remove ☐ save

[Output style](#) ☒ classic ☐ new

PeptGen: output

Classic output:

PeptGen Results

Download a copy of these results in format: [Text](#) [PostScript](#) [PDF](#)

Word length: 18
Overlap consecutive peptides by: 10
Shorten by:
Lengthen by:
Forbidden C-term amino acids:
Forbidden N-term amino acids:
Number of peptides generated: 9
Sequence names: CON_B
CON_C
CON_G

```
HIVWASRELERFAVNPGLLETSEGCRQILGQLQPSLQTGSEELRSYNTVATLYCVHQRIEVKDTKEALEKIEEEQNKSK
HIVWASRELERFAVNPGL (18)
-L-----L-----
-L-----L-----D-
LERFAVNPGLLETSEGCR (18)
----L-----K-----
----L--D-----A--Q-
GLLETSEGCRQILGQLQP (18)
-----K--IK-----
D----A--Q--M-----
CRQILGQLQPSLQTGSEE (18)
-K--IK-----A---T--
-Q--M-----A---T--
QPSLQTGSEELRSYNTV (18)
--A---T-----
--A---T-----F---
EELRSYNTVATLYCVHQ (18)
-----E-----
-----F-----
TVATLYCVHQRIEVKDTK (18)
-----EK---R---
-----
HQRIEVKDTKEALEKIEE (18)
-EK---R-----D----
-----EV-K-----
TKEALEKIEEEQNKSK (16)
-----D-----Q-----
-----EV-KI-K--Q-----
```

Simple output, “new”

```
13 QPSLQTGSEELRSYNTV 5_1.1
14 QPALQTGTEELRSYNTV 5_2.1
15 QPALQTGTEELRSLFNTV 5_3.1

16 EELRSYNTVATLYCVHQ 6_1.1
17 EELRSYNTVATLYCVHE 6_2.1
18 EELRSLFNTVATLYCVHQ 6_3.1

19 TVATLYCVHQRIEVKDTK 7_1&3.2
20 TVATLYCVHEKIEVRDTK 7_2.1

21 HQRIEVKDTKEALEKIEE 8_1.1
22 HEKIEVRDTKEALDKIEE 8_2.1
23 HQRIEVKDTKEALEEVEK 8_3.1

24 TKEALEKIEEEQNKSK 9_1.1
25 TKEALDKIEEEQNKSQ 9_2.1
26 TKEALEEVEKIQKKSQ 9_3.1
```

Distinct

Duplicate

N-Glycosite

- Highlights and tallies predicted N-linked glycosylation sites (Nx[ST] patterns, where x can be any amino acid)
- NP[ST] pattern can be excluded

	210	220	230	240	250	260	270
A1.KE.93.Q23-17	QACPKVSFEP	IPIHYCTPAG	FAILKCKDEG	FNGTGL--CK	NVSTVQCTHG	IKPVVSTQLL	LNGSLAEKNI
B.FR.HXB2	QACPKVSFEP	IPIHYCAPAG	FAILKCNKKT	FNGTGP--CT	NVSTVQCTHG	IRPVVSTQLL	LNGSLAEEEV
C.BR.92.92BR025	QACPKVSFDP	IPIHYCAPAG	YAILKCNKKT	FNGTGP--CN	NVSTIQCTHG	TKPVVSTQLL	LNGSLAEEEI
D.UG.94.94UG1141	QACPKMTFEP	IPIHYCAPAG	FAILKCNEKK	FNGTGP--CK	NVSTVQCTHG	IKPVVSTQLL	LNGSLAEEEI
01_AE.CF.90.90CF11697	QACPKVTFDP	IPIHYCTPAG	YAILKCNEKN	FNGTGP--CK	NVSSVQCTHG	IKPVVSTQLL	LNGSLAEDI
02_AG.CM.97.97CM-MP807	QACPKVSFEP	IPIHFCAPAG	FAILKCKDKE	FNGTGP--CK	NVSTVQCTHG	IKPVVSTQLL	LNGSLAEEKV
CPZ.CM.--.CAM3	QACPKTSFEP	IPIHYCATPG	YAIMKCNMPN	FNGTGTGRCN	NISTVQCTHG	IRPVVTTQLI	LNGSVAENKT
O.CM.--.ANT70	QACPKVSFEP	IPIHYCAPAG	YAIFKCNSTE	FNGTGT--CR	NITVVVCTHG	IRPTVSTQLI	LNGTSLKGKI

N-glycosylation Sites In Each Submitted Sequence

Sequence Name	N-glycosylation Sites Numbers
A1.KE.93.Q23-17	23
B.FR.HXB2	24
C.BR.92.92BR025	23
D.UG.94.94UG1141	24
01_AE.CF.90.90CF11697	24
02_AG.CM.97.97CM-MP807	24
CPZ.CM.--.CAM3	25
O.CM.--.ANT70	25

A total of 192 N-glycosylation sites in 8 sequences have been found.

The total N-Linked glycosylation sites is 192.
(Click the numbers and see the details)

- View the N-Linked glycosylation sites in NX[ST]pattern.
 - The single N-linked glycosylation site count is: 192.
 - The NXS combination count is: [62](#).
 - The NXT combination count is: [130](#).
 - The contiguous N-linked glycosylation site (NN[ST][ST]) count is: 0.

N-Glycosite (continuation)

- Tallies number of N-linked glycosylation sites per alignment position and displays as a downloadable table

Number of N-glycosylation Sites by Position

[Download tab-delimited file](#)

	Position	31	60	69	102	105	109	110	111	112	115	122	128	145	149	174	179	182	184	185	195	227
A1.KE.93.Q23-17			N			N				N				N	N							N
B.FR.HXB2			N					N				N		N	N		N					N
C.BR.92.92BR025			N			N						N		N	N		N		N			N
D.UG.94.94UG1141			N		N							N		N	N		N			N	N	
01_AE.CF.90.90CF11697			N						N			N	N	N	N		N			N	N	
02_AG.CM.97.97CM-MP807			N				N				N	N		N	N					N	N	
CPZ.CM.-.CAM3			N	N					N			N		N	N		N					N
O.CM.-.ANT70		N	N												N	N		N			N	N
Total		1	8	1	1	2	1	1	2	1	1	6	1	7	8	1	5	1	1	3	8	1

ELF

- If you have a peptide that reacts with CD8+ T cells from a person with known HLA type, enter:
 - ☐ The peptide that reacts with CD8+ T-cells
 - ☐ The HLA type of the person with the reactive CD8+ T cells

- ELF will help identify the possibly reactive epitope by
 - ☐ Highlighting appropriate HLA anchor motifs in the peptide
 - ☐ Aligning all known epitopes embedded in the peptide from the database to your query sequence, with links to epitope entries
 - ☐ Finding potential epitopes based on Immune Epitope Database (IEDB) binding predictions <http://www.immuneepitope.org/>

- Other useful information provided:
 - ☐ Genomic location of your peptide
 - ☐ Database records for known CTL epitopes in this region, regardless of HLA.

ELF

Epitope Location Finder

Purpose: search a submitted protein sequence for (1) known epitopes from our immunology databases, (2) epitopes predicted by consensus binding motifs, and (3) epitopes predicted by the IEDB binding algorithm. For details see [ELF Explanation](#).

Input

Paste [protein sequence](#) DTVLEDMNLPGRWKPKMIG <50 amino acids, raw format

Options

Show [known epitopes](#) ☒ from CTL and Helper databases

Find potential epitopes ☒ based on [anchor residues](#)

Choose [HLA\(s\)](#)
(Class I and Class II)

Use control-click for multiple selection

By genotype

A*3004
A*3101
A*3201
A*3303
A*6601
A*6801
A*6802

By serotype

A33(19)
A69(28)
A68(28)
A30(19)
A66(10)
A1
A2

Find potential epitopes ☒ based on [IEDB binding predictions](#)

Choose [HLA\(s\) or MHC\(s\)](#)
(synchronized with genotype selections above)

HLA Class I

A*6611
A*6612
A*6613
A*6614
A*6615
A*6801
A*6802

HLA Class II

DRB3*0224
DRB3*0225
DRB3*0301
DRB3*0303
DRB4*0101
DRB4*0103
DRB5*0101

Animal MHC Class I

chimpanzee
Patr-A*0101
Patr-A*0201
Patr-A*0301
Patr-A*0302
Patr-A*0401
Patr-A*0402

Animal MHC Class II

mouse
H2-IAb
H2-IAd
H2-IEd

Display binders ☒ Show best binder(s) per MHC

☐ Show below [percentile rank](#) (1-100) per MHC

E-mail result ☐ Predictions are slow. For more than a few HLAs/MHCs, we recommend e-mailed result.

Submit


Reset

HLA selection is synchronized between 2 analysis options

You can choose how many top binders to show per MHC, or use a binding percentile rank cutoff

ELF results 1:


Epitopes from our CTL database aligned to your query sequence


Bold **red** letters indicate residues that differ from the query sequence. The symbol  means the HLA of the epitope matches one of your submitted HLAs. Click on the epitope to see full database entry. Click on "align" to align the epitope to the sequence database via QuickAlign.


Epitopes shown here are completely within the bounds of your query. Epitopes that overlap the ends of your query are included in the "View database records" links above.

[Download](#) this alignment in format table

DTVLEDMNLPGRWKPKMIG

[DTVLE**EM**NL](#) A*6802 [align](#) 

[DTVLE**I**NL](#) A*6802 [align](#) 

[DTVLE**EW**NL](#) A*6802 [align](#) 

[DTVLE**EM**NL](#) A68 [align](#)

[DTVLE**EM**NL](#) A28 [align](#)

[DTVLEDMNL](#) [align](#)

[E**EM**NLPGRW](#) B44 [align](#)

[E**E**I**N**LPG**KW**](#) B44 [align](#)

[E**EM**NLPGRW](#) B*4402 [align](#)

[E**EM**NLPGRW](#) B*4403 [align](#)

[E**EM**NLPGRW](#) B18,B40,B44 [align](#)

[EDMNLPGRW](#) [align](#)

[E**EM**NLPGRW](#) B*44 [align](#)

[E**E**I**N**LPG**KW**](#) B*4403 [align](#)

[E**EM**NLPGRW](#) [align](#)

[LPGRWKPKMI](#) Cw3 [align](#)

[LPGRWKPKMI](#) B7 [align](#)

Clicking on an epitope takes you to respective CTL or Helper epitope Database entries

ELF results 2:

Potential epitopes based on anchor residues

These peptides have C-terminal anchor residues, highlighted in **blue**, and internal anchors highlighted in **magenta**. These anchor residues match one or more motifs associated with the submitted HLA.

[Download](#) this alignment in format table

DTVLEDMNLPGRWKPKMIG

DTVLEDM**N**L (A*0205[L])

D**T**VLEDM**N**L (A*6802 ..[TV].....[VL])

TVLEDMNLP (A*0206 ..[VQ].....)

LED**M**NLP**G**R (DRB5*0101,DRB5*0101 [FYLM]..[QVIM]....[RK])

ELF results 3:

Potential epitopes based on IEDB database MHC binding predictions, by Alexander Sette's group

Potential epitopes based on IEDB binding predictions

Top binders for each MHC are highlighted in [blue](#).

Prediction method: IEDB recommended

Low percentile = good binders

Show up to 1 binder(s) per MHC

Class I

Selected allele(s): A*6802, B*1501

this alignment in format

DTVLEDMNLPGRWKPKMIG (Click MHC to see full list of IEDB predictions for that MHC)

[DMNLPGRW](#) [B*1501](#) (26)

[MNLPGRWK](#) [A*6802](#) (3.0)

Clicking on MHC links to the full list of IEDB predictions for that MHC (see next slide)

Class II

Selected allele(s): DRB5*0101

this alignment in format

DTVLEDMNLPGRWKPKMIG (Click MHC to see full list of IEDB predictions for that MHC)

[TVLEDMNLPGRWKPK](#) [DRB5*0101](#) (17.17)

ELF results 3:

Potential epitopes based on IEDB database MHC binding predictions, by Alexander Sette's group

IEDB Analysis Resource

[Home](#)[Help](#)[Example](#)[Reference](#)[Download](#)[Contact](#)

MHC-I binding predictions - Prediction Results

Input Sequences

#	Name	Sequence
1	sequence 1	DTVLEDMNLPGRWKPKMIG

Prediction method: IEDB recommended | Low percentile = good binders

Check to expanded the result: ☐

Allele	#	Start	End	Peptide Length	Sequence	Method used	Percentile Rank
HLA-B*15:01	1	6	13	8	DMNLPGRW	NetMHCpan	26
HLA-B*15:01	1	3	13	11	VLEDMNLPGRW	NetMHCpan	27
HLA-B*15:01	1	3	11	9	VLEDMNLPG	Consensus (ANN,SMM,CombLib_Sidney2008)	27.60
HLA-B*15:01	1	8	17	10	NLPGRWKPKM	NetMHCpan	31
HLA-B*15:01	1	7	17	11	MNLPGRWKPKM	NetMHCpan	35
HLA-B*15:01	1	2	9	8	TVLEDMNL	NetMHCpan	36
HLA-B*15:01	1	2	11	10	TVLEDMNLPG	NetMHCpan	47
HLA-B*15:01	1	4	11	8	LEDMNLPG	NetMHCpan	48

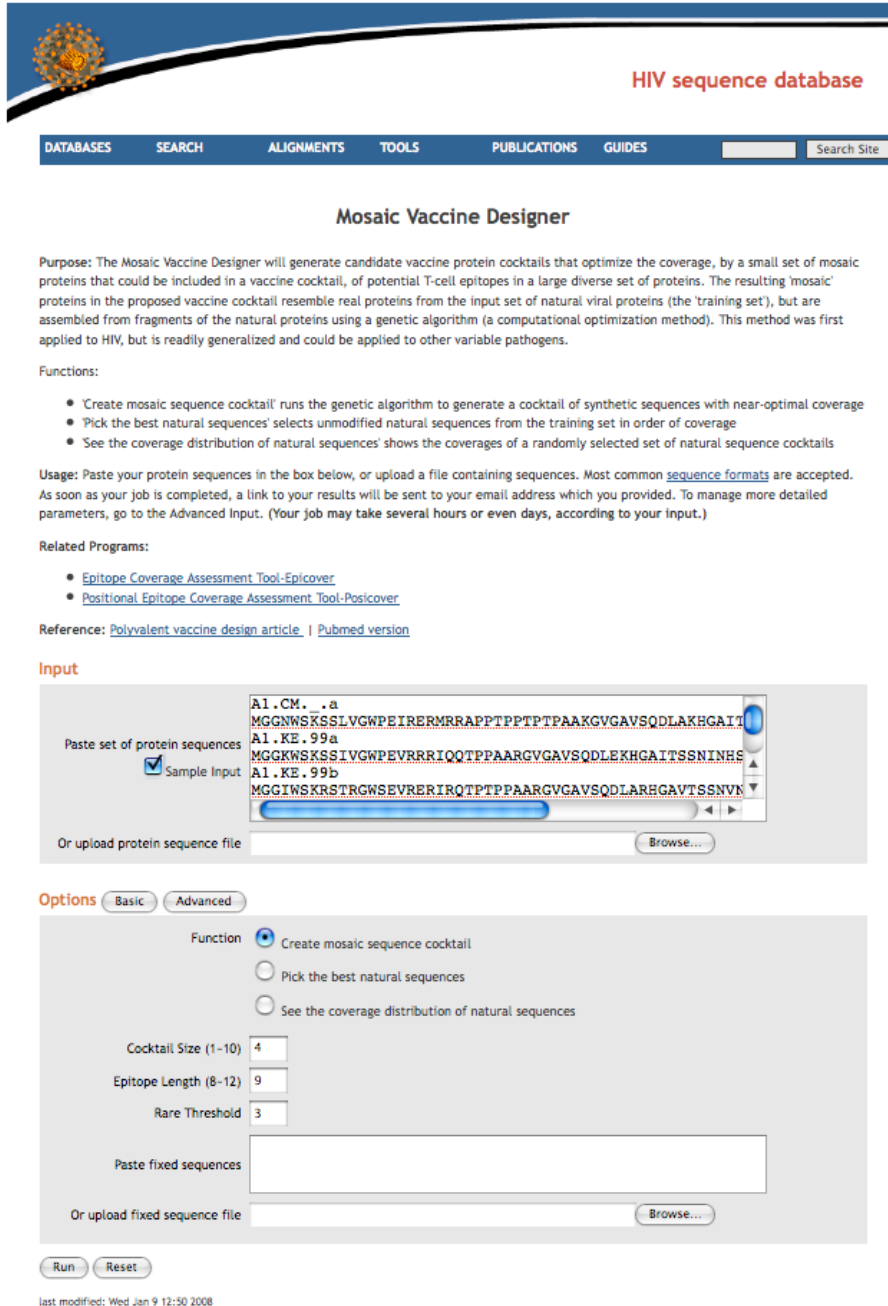
Mosaic vaccine tools

Mosaic Vaccine Designer: The Mosaic Vaccine Designer will generate candidate vaccine protein 'cocktails' that optimize coverage of potential T-cell epitopes found in a given background set of protein sequences.

Epitope Coverage Assessment: Alignment independent “n-mer” coverage of sequences by vaccines or peptides.

Positional Epitope Coverage Assessment: Alignment dependent coverage of sequences by vaccines or peptides.

Mosaic Vaccine Designer



The image shows the Mosaic Vaccine Designer web application interface. At the top, there is a header with a logo on the left and the text "HIV sequence database" on the right. Below the header is a navigation bar with links: DATABASES, SEARCH, ALIGNMENTS, TOOLS, PUBLICATIONS, GUIDES, and a search box. The main content area is titled "Mosaic Vaccine Designer". It contains a "Purpose" section explaining the tool's function, a "Functions" section with three bullet points, a "Usage" section with instructions, and a "Related Programs" section with two links. Below this is a "Reference" section with two links. The "Input" section has a text area for pasting protein sequences, a "Sample Input" checkbox, and a "Browse..." button for uploading a file. The "Options" section has two tabs: "Basic" and "Advanced". Under "Basic", there are three radio buttons for the function: "Create mosaic sequence cocktail" (selected), "Pick the best natural sequences", and "See the coverage distribution of natural sequences". There are also input fields for "Cocktail Size (1-10)" (set to 4), "Epitope Length (8-12)" (set to 9), and "Rare Threshold" (set to 3). There is a "Paste fixed sequences" text area and a "Browse..." button for uploading a fixed sequence file. At the bottom, there are "Run" and "Reset" buttons. A footer note states "last modified: Wed Jan 9 12:50 2008".

HIV sequence database

DATABASES SEARCH ALIGNMENTS TOOLS PUBLICATIONS GUIDES Search Site

Mosaic Vaccine Designer

Purpose: The Mosaic Vaccine Designer will generate candidate vaccine protein cocktails that optimize the coverage, by a small set of mosaic proteins that could be included in a vaccine cocktail, of potential T-cell epitopes in a large diverse set of proteins. The resulting 'mosaic' proteins in the proposed vaccine cocktail resemble real proteins from the input set of natural viral proteins (the 'training set'), but are assembled from fragments of the natural proteins using a genetic algorithm (a computational optimization method). This method was first applied to HIV, but is readily generalized and could be applied to other variable pathogens.

Functions:

- 'Create mosaic sequence cocktail' runs the genetic algorithm to generate a cocktail of synthetic sequences with near-optimal coverage
- 'Pick the best natural sequences' selects unmodified natural sequences from the training set in order of coverage
- 'See the coverage distribution of natural sequences' shows the coverages of a randomly selected set of natural sequence cocktails

Usage: Paste your protein sequences in the box below, or upload a file containing sequences. Most common [sequence formats](#) are accepted. As soon as your job is completed, a link to your results will be sent to your email address which you provided. To manage more detailed parameters, go to the Advanced Input. (Your job may take several hours or even days, according to your input.)

Related Programs:

- [Epitope Coverage Assessment Tool-Epicover](#)
- [Positional Epitope Coverage Assessment Tool-Posicover](#)

Reference: [Polyvalent vaccine design article](#) | [Pubmed version](#)

Input

Paste set of protein sequences

☒ Sample Input

Or upload protein sequence file

Options

☒ Basic ☐ Advanced

Function

☒ Create mosaic sequence cocktail

☐ Pick the best natural sequences

☐ See the coverage distribution of natural sequences

Cocktail Size (1-10)

Epitope Length (8-12)

Rare Threshold

Paste fixed sequences

Or upload fixed sequence file

last modified: Wed Jan 9 12:50 2008

Input: protein sequence set for a target population, does not need to be aligned.

Number of mosaic proteins in the set.

Epitope length.

Epitope Coverage Assessment - Epicover

Input

Use output from MakeVaccine tool

Provide a job number to access output from the [Mosaic Vaccine Designer](#) tool:

OR

Provide input sequences

Paste antigen protein
sequence(s):
[\[Sample Input\]](#)

upload more [+] antigen sequence files

and/or upload as files:

Browse...

Paste test set protein
sequences:

upload more [+] test sequence files

and/or upload as files:

Browse...

Options

Send results as an email instead of displaying in browser
(useful in case of a browser time-out): ☐

Nominal epitope length:

Maximum amino acid mismatches to score (range from 0):

Minimum number of occurrences of a potential epitope
in viral protein set to consider for coverage:

Precision to use when reporting coverage: decimal places

Advanced Options

Upload file of grouped sequence names

Browse...

Report on subsets defined according to first character(s) in sequence names

Submit

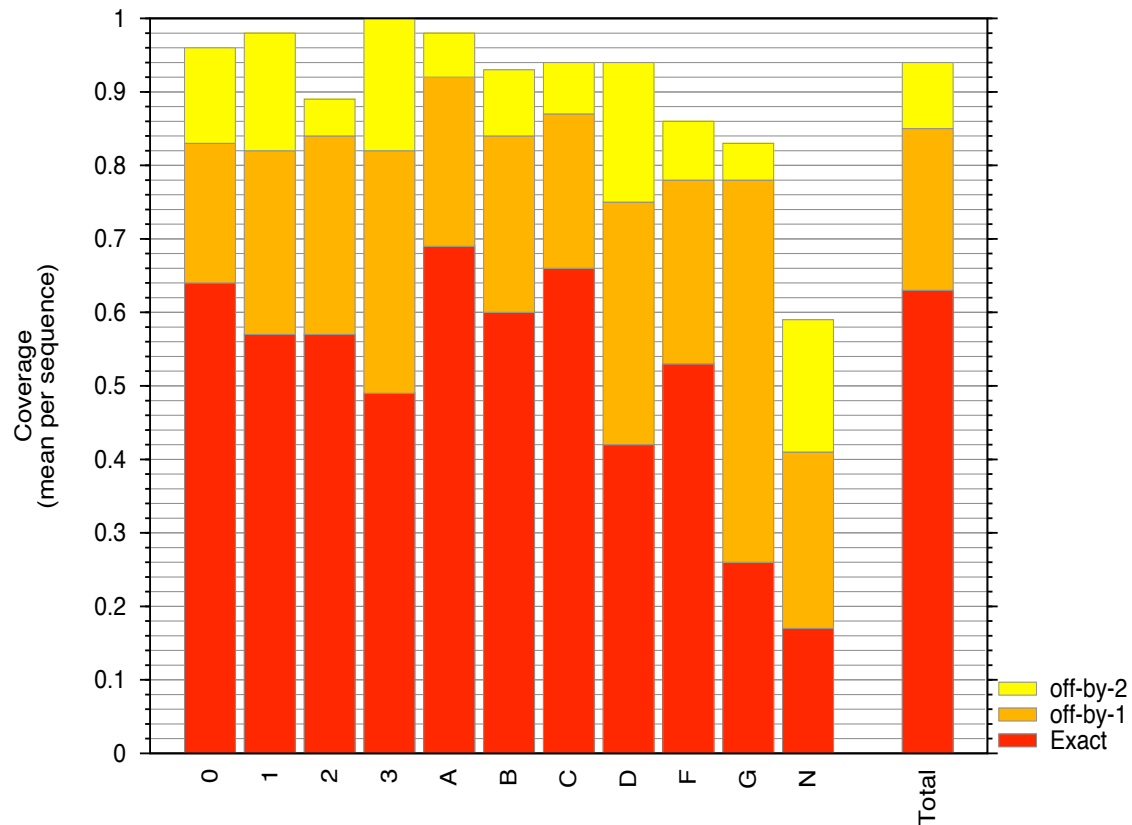
Reset

Input:
Vaccine set
Test set

Can report on
subsets defined
according to the
first several
characters in
sequence
names or
user-defined
subsets

Epicover output

vaccine set	subset	subset count	Off-by-0	Off-by-1	Off-by-2	rare(<3,>1)	unique	absent
vaccine_set_from_user	Total	63	0.6615	0.8914	0.9660	104	114	334
vaccine_set_from_user	A	11	0.7232	0.9429	0.9935	47	36	334
vaccine_set_from_user	B	11	0.6378	0.8845	0.9755	25	19	334
vaccine_set_from_user	C	35	0.6921	0.8994	0.9637	51	45	334
vaccine_set_from_user	D	4	0.4217	0.7546	0.9443	4	9	334
vaccine_set_from_user	F	1	0.5300	0.7800	0.8600	4	5	334
vaccine_set_from_user	G	1	0.2597	0.7792	0.8312	0	0	334



Positional Epitope Coverage Assessment - Posicover

Provide a job # from
Mosaic Vaccine Designer: (Only the antigen set is used. Provide the ALIGNED viral test set below)

AND/OR

Paste antigen protein set
or peptide cocktail:
(alignment not required)
[\[Sample Input\]](#)

upload more [+] antigen files

and/or upload antigen
file(s):

Input:

Vaccine set or
Reagent set

Test set proteins

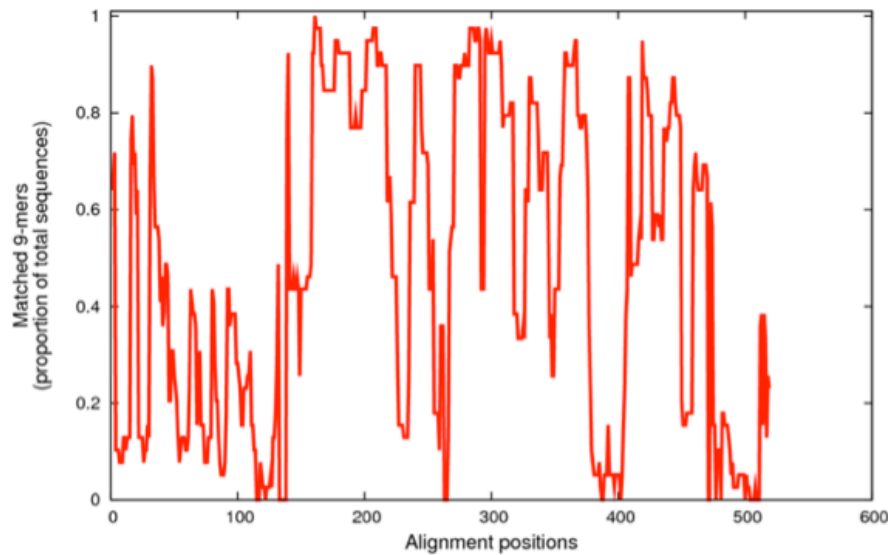
Paste **ALIGNED** test viral
protein set:
[\[Sample Input\]](#)

or upload an **ALIGNED** test
proteins file:

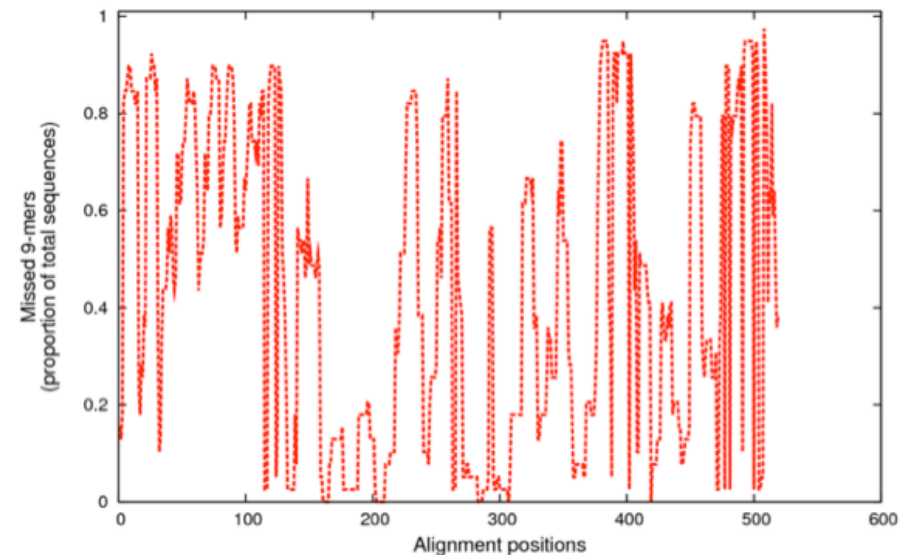
ALIGNED test
set

Examples of Posicover outputs

Matched 9-mers

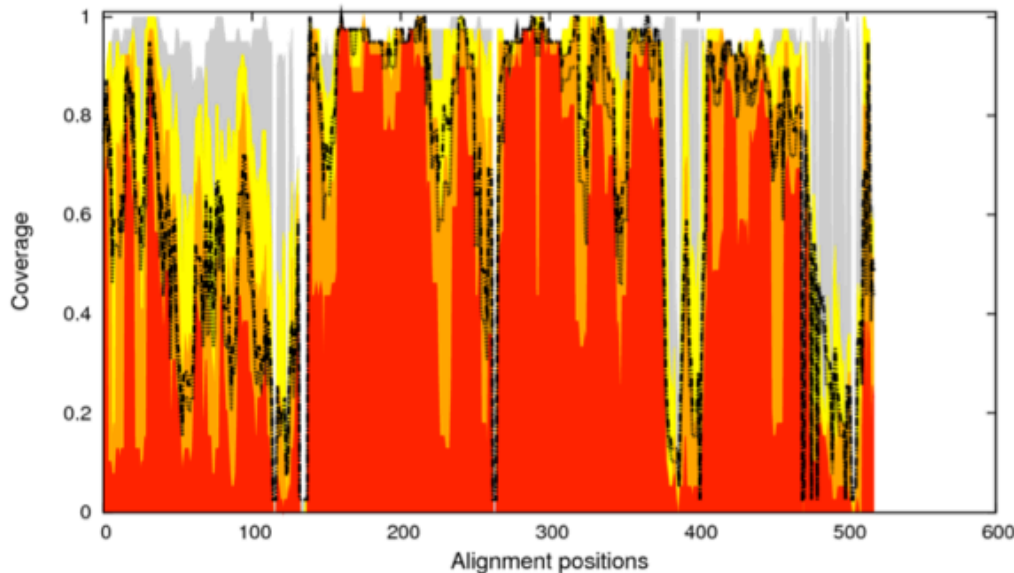


Missed 9-mers



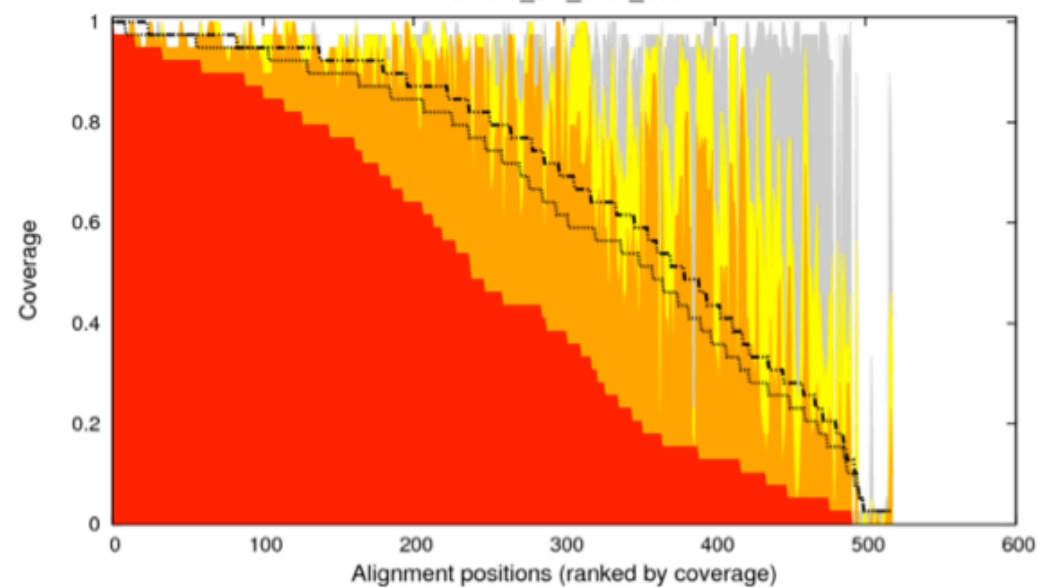
Alignment positions

9-mer coverage by position vaccine_set_from_user



total 9-mers █ exact match █
 up to 7/9 match █ Upper bound: 3 antigen(s) ---
 up to 8/9 match █ Upper bound: 4 antigen(s) -.-.-

Ranked 9-mer coverage vaccine_set_from_user



total 9-mers █ exact match █
 up to 7/9 match █ Upper bound: 3 antigen(s) ---
 up to 8/9 match █ Upper bound: 4 antigen(s) -.-.-

Examples of Posicover outputs

User's sequence alignment:

Each aa is represented as a single-colored square



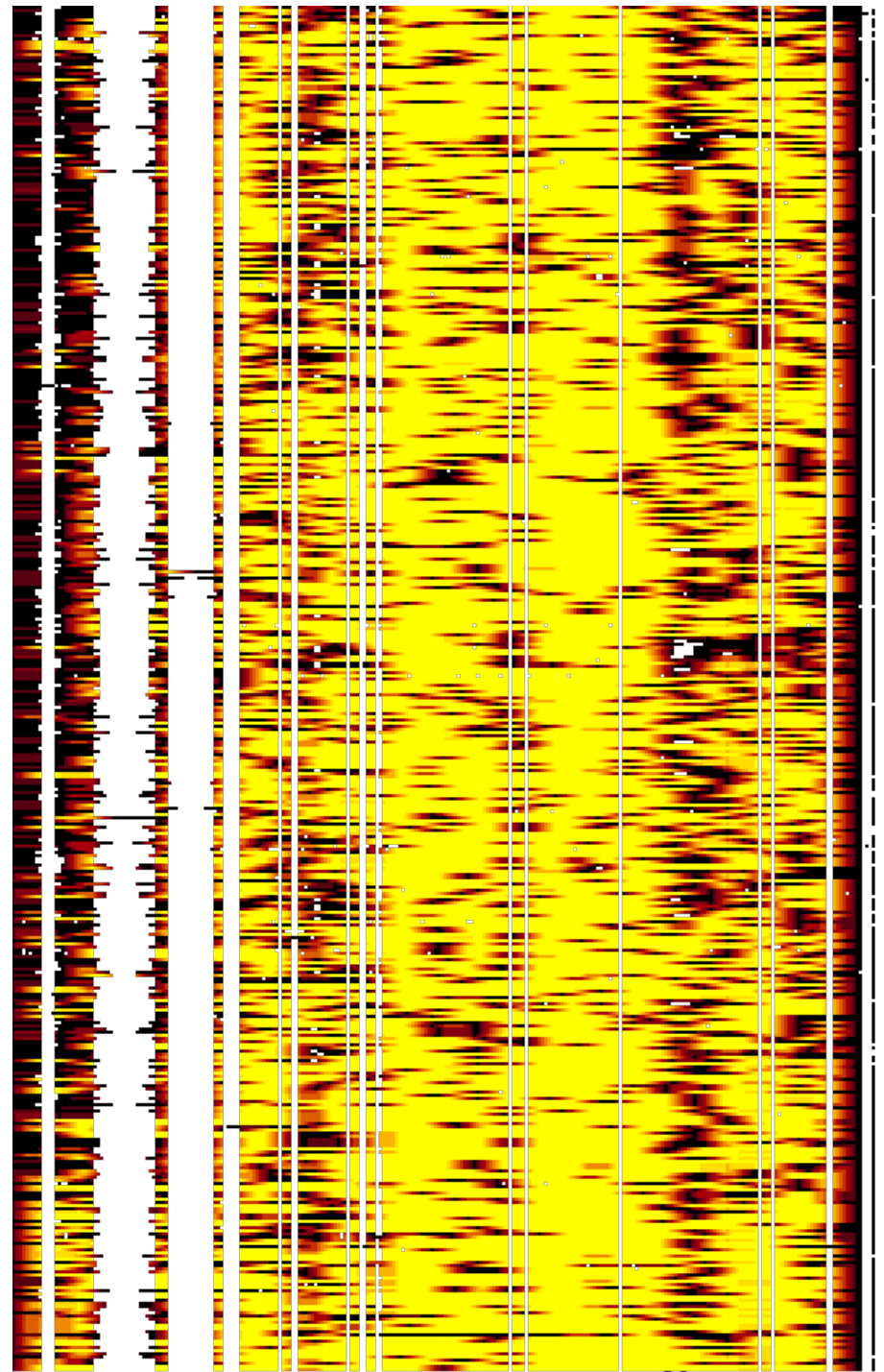
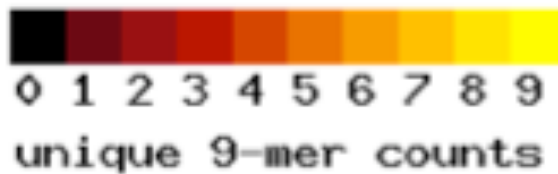
Examples of Posicover outputs

Each amino acid is colored according to the set of 9-mers that contain it:

Yellow: all 9-mers that overlap with amino acid are perfectly matched in a test vaccine set;

Increasingly red: fewer and fewer matches in the overlapping set of 9-mers that span the amino acid;

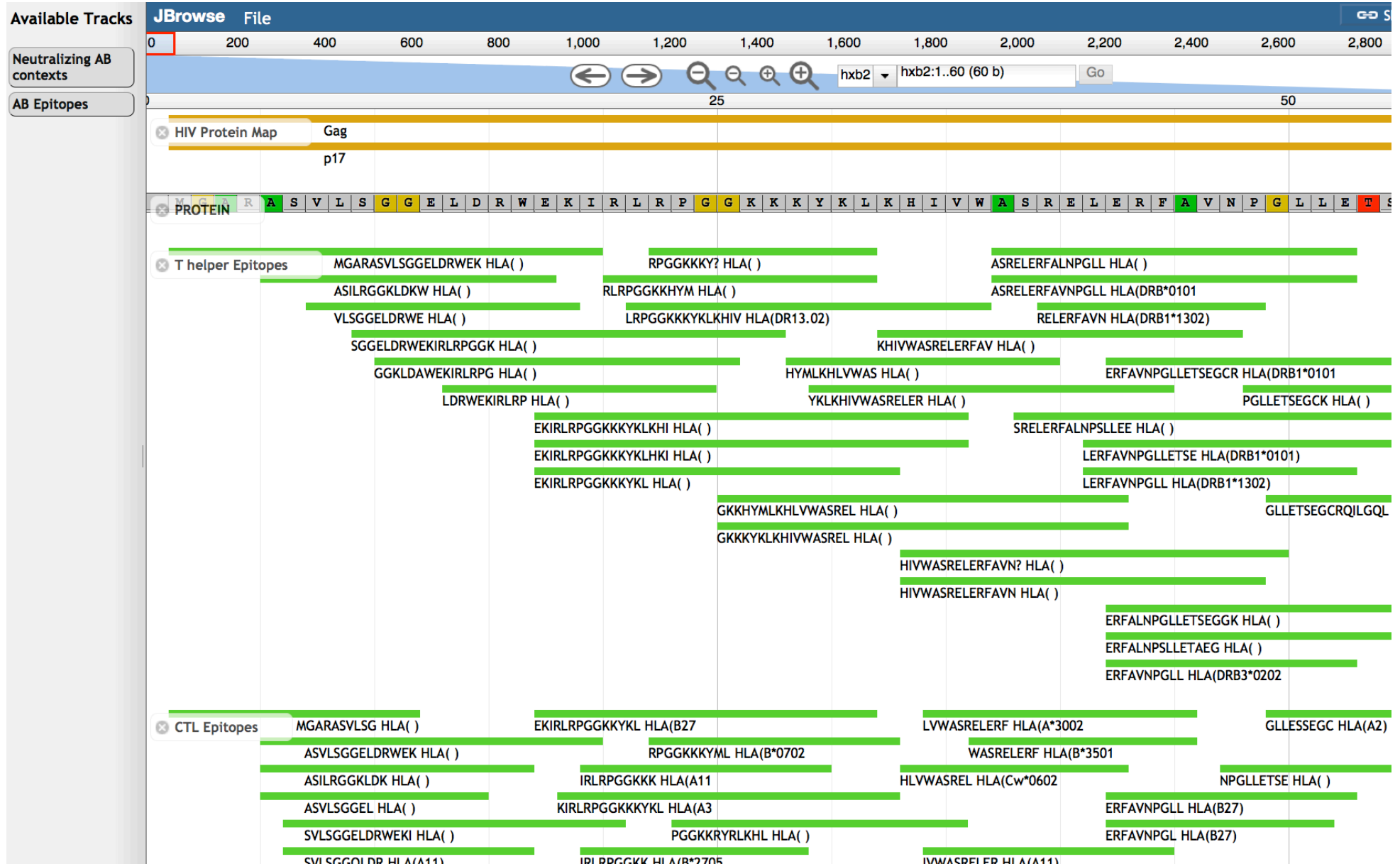
Black: amino-acid residues that are not included in any vaccine set



Coming soon: HIV-1 genome browser



Coming soon: HIV-1 genome browser



**Please let us know if you have
questions, comments or
suggestions**

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